

Antimicrobial activity of vaginal Lactobacillus against urogenital pathogensSuhad Faisal Hatem Al-Mugdadi^{1*}, Ishraq Hasan Elewi, Zainab Farooq Shafeeq Al-Ryyis, Hala Muaed, Mays Ali**Abstract**

Lactobacillus is one of the most important genera of Lactic Acid Bacteria (LAB). Lactobacilli present in a healthy vagina are part of normal bacterial microbiota and protect the host from urogenital infections by maintaining a low pH. Important features of probiotic lactobacilli are to achieve antagonistic activity against bacterial pathogens because of their capacity to produce lactic acid and other organic acids, H₂O₂ and bacteriocin. This study aimed to studying the effect of Lactobacillus spp. isolated from healthy vagina as a probiotic against many clinical bacterial isolates from urogenital tract infection. Susceptibility to Five antibiotics, such as amikacin (AK), Tobramycin (TOB), Piperacillin (PRL), Nalidixic acid (NA) and Gentamycin (GN) was achieved on all pathogenic bacteria isolated from urine and vagina. Two-fold dilutions of Lactobacillus sp. were prepared (1×10^8 , 1×10^4 and 1×10^2) CFU/ml to determine antibacterial activity against urogenital bacteria isolates using agar well diffusion method. N. gonorrhoeae from vagina was affected by all these dilutions of Lactobacillus sp. with inhibition zones 10, 8 and 8 mm respectively. While inhibition zones of S. aureus and Candida albicans from vagina were 12mm in dilution (1×10^4) and 6 mm in dilution (10^8) respectively. In addition, there was no biological activity of Lactobacillus sp. against bacteria isolated from urine. The effectiveness of natural lactobacilli from vagina consider as good therapeutics against bacterial illness of humans. We need future study to purification the active component like bacteriocin or other organic acids from these bacteria.

Keywords: Antibiotics; lactobacilli, vagina; Probiotic; Agar well diffusion method* Correspondence author: suhadhatim@uomustansiriyah.edu.iqSuhad Faisal Hatem Al-Mugdadi¹, orcid.org/0000-0002-7164-7093Ishraq Hasan Elewi, orcid.org/0000-0002-7918-7853Zainab Farooq Shafeeq Al-Ryyis, orcid.org/0000-0001-6514-7925¹University of Al-Mustansiriyah/ College of Pharmacy/Department of Clinical Laboratories sciences

Received 12 August 2017, Accepted 30 November 2017, Available online 27 December 2017

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Introduction

Lactobacillus is one of the most important genera of Lactic Acid Bacteria [1]. Lactic acid bacteria (LAB) are Gram positive, non-spore forming catalase negative cocci or fermentative lactobacilli which produce lactic acid from fermentation of carbohydrates [2,3], facultative anaerobic, rod shape and non-spore forming bacteria [4]. The Genus Lactobacillus currently includes more than 170 different species [5] and only a part of them are representatives of protective vaginal microbiota [6]. Also, lactobacilli can participate in the protection of the vaginal epithelium by the means of several other barrier mechanism Lactobacillus could be used as prophylactic and therapeutic agents [7]. Lactobacilli present in a healthy vagina are part of normal bacterial microbiota and protect the host from urogenital infections by maintaining a low pH (<4.5), by producing bacteriostatic and bactericidal substances and through competitive exclusion [8,9]. Are excellent antibacterial agents (major part of the vaginal micro biota [10,11]. Also, lactobacilli can participate in the protection of the vaginal epithelium by the means of a number of other barrier mechanisms Lactobacillus could be used as prophylactic and therapeutic agents [7]. One of the important features of probiotic lactobacilli is to achieve antagonistic activity against bacterial pathogens because of their capacity to produce lactic acid and other organic acids that lower the pH in the human intestine, and to produce H₂O₂ and bacteriocin, thereby establishing a hostile environment for the growth and survival of various human pathogenic bacteria [12,13]. Studied the effect L.acidophilus strains in combination with antibiotics, and found that

the strains improved the antibacterial efficacy of the test antibiotics against E. coli, Salmonella typhimurium, S. aureus and Shigella flexeneri. However, no report has been made on the combined antibacterial activity of two different lactobacilli strains, at least in this part of the globe, against human pathogenic bacteria Probiotics are defined as live microorganisms which when administered in adequate quantity confer health benefits to the host and lactobacilli of human origin are potential probiotics against urogenital tract infections [14, 15]. Many isolated Lactobacilli were examined against multi-drug-resistant uropathogens, including E. coli, C. albicans, E. fecalis, P. aeruginosa and K. pneumoniae and showed a good results as antimicrobial agent [16]. Bacterial vaginosis and vulvovaginal candidiasis are the most common vaginal infections worldwide [17]. The first one is responsible for about 50% of all the cases and significantly associated with reduce level in lactobacilli population and increase in anaerobic and facultative aerobic pathogens [18, 19]. The second one affects about 75% of women with usual complaints like pruritus and vaginal discharge [20]. The majority of vulvovaginal candidiasis cases (90%) caused by Candida albicans [21, 22]. Among the Eleven vaginal lactobacilli isolates, L. agilis, L. jensenii, L. johnsonii, and L. ruminus acted as potential probiotic species including the ability to co-aggregate with Candida species, adhere to epithelial mucosa, to auto-aggregate, and to create both H₂O₂ and lactic acid. Although H₂O₂ production by all strains, the highest levels were noticed for L. acidophilus, L. gasserii, L. crispatus, L. vaginalis and L. johnsonii [23]. This study aimed to studying the effect of Lactobacillus spp. isolated from healthy vagina as a probiotic against many clinical

bacterial isolates from urogenital tract infection which are multidrug resistance.

Materials and Methods

Isolation and characterization of lactobacillus spp.

The lactic acid bacteria were isolated from healthy vaginal swabs on selective media. This isolate were cultured and stored in De Man Rogosa and Sharpe (MRS) media (Hi-Media, India) on agar plate or broth incubated anaerobic (5-10%) in candle jar for a period of 18-36h h at 37 °C [24]. From MRS agar plates, the diagnosis of lactobacilli sp. depended on morphology (gram positive rod-shaped bacteria), culture, non-motile, non-spore forming, negative oxidase and catalase test.

Clinical isolates

All clinical isolates used in this study were collected from patients suffered from urogenital clinical sign and symptoms and attended Bagdad teaching hospital-Iraq. All samples plated on agar media including Blood, Macconkey and Chocolate agar which were prepared according to the instructions of the Hi- media –India company. at 37°C for 18-24h. The most pathogenic bacteria were diagnosed by microscope examination, culture and some biochemical tests. Eight pathogens submitted in this study included: Four kinds of pathogens (K. pneumonia, E. coli, M. morgani and S. aureus) isolated from urine samples, also another four samples (N. gonorrhoeae, S. aureus, Proteus spp. and Candida albicans) isolated from vagina.

Susceptibility pattern of clinical isolates

Susceptibility to 5 antibiotics, such as amikacin (AK), Tobramycin (TOB),

Pipracillin (PRL), Nalidixic acid (NA) and Gentamycin (GN) (Hi-Media,India) were tested against urogenital pathogens by using disc diffusion method [25]. Single colonies of the test bacteria strains were inoculated into Muller Hinton agar and incubated at 37°C for 18-24 h. by cotton swab which was used to spread bacteria on the surface, and the antibiotic discs were placed on the surface of the agar plates. After incubation for 24 h at 37°C, the zone diameter of inhibition (ZDI) obtained around the antibiotic disc were recorded.

Fresh Lactobacillus spp. dilutions

Two-fold dilutions were prepared from lactobacillus sp. include (10^8 , 10^4 and 10^2) CFU/ml adjusting to McFarland to determine the effect all these dilutions against the pathogenic bacteria. 100 ml of inoculums of each type of pathogenic bacteria was spread on muller- Hinton agar plate and blood agar. Then let the plate to dry then, six mm wells were made in agar plates. 100µl volume of each dilution (10^8 , 10^4 and 10^2) submitted in wells into media. All plates were incubated at (37°C for 18-36h.). Antimicrobial activity determined by clear inhibition zone surrounding the wells, results consider positive if the inhibition zone more than 6mm. completely inhibited the growth for 24h was recorded.

Results

Table (1) showed multidrug resistant bacteria against five antibiotic types including disc of Amikacin (AK), Tobramycin (TOB), Pipracillin (PRL), Nalidixic acid (NA) and Gentamycin (GN). S. aureus isolated from urine and vagina in addition to N. gonorrhoeae from vagina were resisted to all five antibiotic types. K. pneumonia was sensitive to Amikacin and

Gentamycin and resist to others. *Proteus* spp. sensitive to Amikacin and Nalidixic acid and resist to others. Two-fold dilutions of natural lactobacillus sp. Isolate from healthy vagina were prepared as following (1×10^8 , 1×10^4 and 1×10^2) *N. gonorrhoeae* from vagina was affected by all these dilutions with inhibition zones 10, 8 and 8

mm respectively. While inhibition zones of *S. aureus* and *Candida albicans* from vagina were 12 in dilution (1×10^4) and 6 mm in dilution (1×10^8) respectively. Bacteria isolated from urine (*K. pneumonia*, *E. coli*, *M. morganii* and *S. aureus*) were resistant to all dilutions of lactobacillus sp., Table (2) and Figure (1).

Table 1

Sensitivity pattern of some antibiotic disk against many clinical bacterial isolates

Antibiotic disk Bacterial isolates	AK	GN	PRL	NA	TOB
<i>K. pneumonia</i> (urine)	S	S	R	R	R
<i>E. coli</i> (urine)	S	R	R	S	R
<i>M. morganii</i> (urine)	S	R	R	R	R
<i>S. aureus</i> (urine)	R	R	R	R	R
<i>Proteus</i> spp.(vaginal)	R	R	R	S	R
<i>S. aureus</i> (vagina)	S	R	R	R	R
<i>N. gonorrhoeae</i> (vagaina)	R	R	R	R	R
<i>Candida albicans</i> (vagaina)	-	-	-	-	-

Table 2

In vitro activity of newly isolated vaginal lactobacillus sp. dilutions against many pathogens

Lactobacillus sp. dilutions Bacterial isolates	Diameter of inhibition zone (mm)		
	1×10^8	1×10^4	1×10^2
<i>K. pneumonia</i> (urine)	-----	-----	-----
<i>E.coli</i> (urine)	-----	-----	-----
<i>M. morganii</i> (urine)	-----	-----	-----
<i>S. aureus</i> (urine)	-----	-----	-----
<i>Proteus</i> spp. (vaginal)	-----	-----	-----
<i>S. aureus</i> (vagina)	-----	12mm	-----
<i>N. gonorrhoeae</i> (vagaina)	10mm	8mm	8mm
<i>Candida albicans</i> (vagaina)	6mm	-----	-----

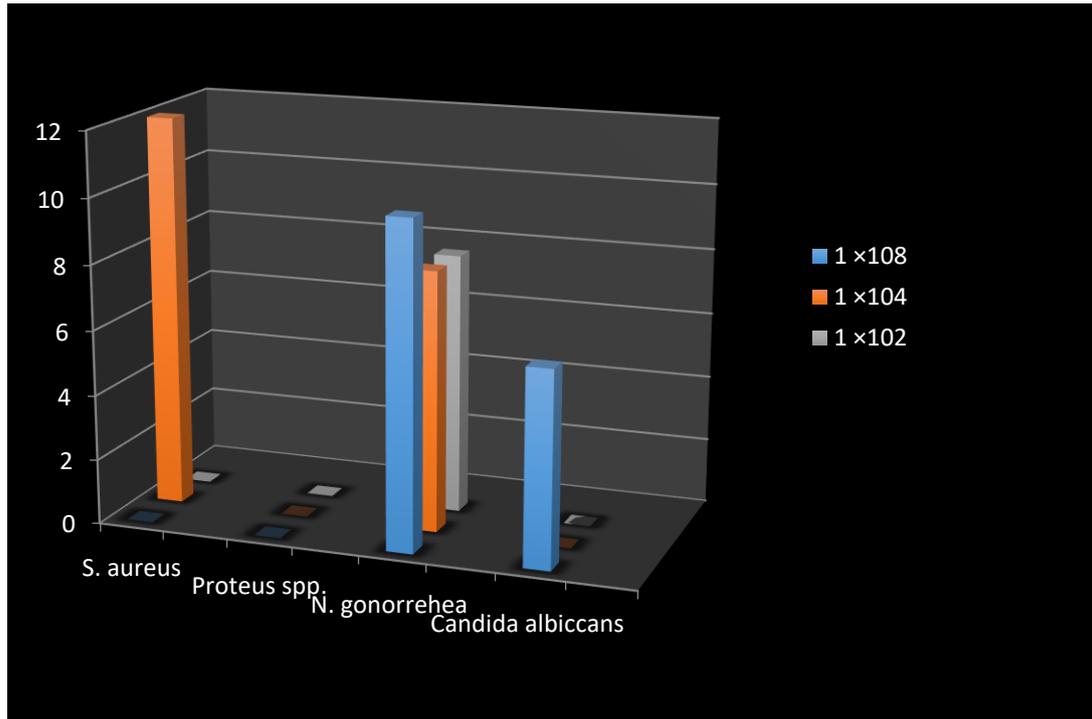


Figure 1

lactobacillus sp. dilutions activity against four kinds pathogens isolated from vagina

Discussion

This increasing in the emergence of drug-resistant pathogenic bacteria led to search the non-antibiotic treatment regimens, including the probiotic lactobacilli, as new therapies to fight the bacterial antibiotic resistance and to treat the disease [26-27]. The result in Table (1) showed resistant of bacteria to word Five antibiotics namely mikacin; Pipracillin; Gentamycin; Tobramycin and Nalidixic acid. Antibiotic resistant infections can occur anywhere in the community but also there has been an increase of infections that occur in the hospitals and other medical facilities.

Resistance in bacteria is a large issue because as bacteria evolves and forms ways to make the antibiotics ineffective and harmful and fatal bacteria can thrive in multiple environments [28]. [29]. Cited that Strains of E. coli that are resistant to single or multiple classes of antibiotics, also pneumococcal resistance has increased to Beta-lactams, Macrolides and Lincosamides while M. Morgani & Proteus spp. isolates resistant to third-generation cephalosporins, imipenem, amikacin, and ticarcillin. As we know that bacteria resist to multi drug when it carries several resistance genes. It should be noted that a relatively high biological activity of half dilutions of Lactobacillus sp.

isolate isolated from healthy vagina (1×10^8 , 1×10^4 , 1×10^2) against *S. aureus*, *N. gonorrhoeae* and *Candida albicans* isolates from vagina by measuring the inhibitory effect zone (mm) as shown in table(2), this result lined with [30,23]. The *Lactobacillus* species are usually selected as the probiotic microorganisms because of and bile, inhibitory effect against pathogenic microbes, and innate resistance to antibiotics. The production of lactic acid and other organic acids, hydrogen peroxide (H_2O_2) and bacteriocins help to keep the vaginal pH below 4.5 and creates a hostile ecosystem for the growth and survival of pathogenic microorganisms [23,31]. However, it did not inhibit other bacteria isolate in current study may be because the inhibitory activity of this bacteria (antagonistic effect) occurred between pH 3 and pH 5 which it not adjusted in vitro. [32] Reported that several antifungal compounds, such as cyclic dipeptides, pyroglutamic acid and lactones were formed by *Lactobacillus* isolates and played an important role against *Candida* spp. in addition to ability of probiotic strains to adhere to epithelial mucosa, to auto-aggregate, co-aggregate with *Candida* species, and to produce both H_2O_2 and lactic acid. All the lactic acid bacteria screened for bacteriocin production, *Lactobacillus bulgaricus*, *L. lactis*, *L.*

acidophilus, *Lactococcus lactis*, *Streptococcus thermophilus*, *S. cremoris*, *Pediococcus halophilus* and *P. cerevisiae* produced bacteriocin activity among 4800 and 6000 Au/ml against *Staphylococcus*, *Salmonella*, *Shigella*, *Bacillus* and *Pseudomonas* species [33]. The susceptibilities of the examined Gram-positive (*S. aureus*, *S. xylosus* and *S. uberis*) and Gram-negative bacteria (*E. coli* and *Y. enterocolitica*) to growth inhibition by the bacteriocin of *Lactobacillus* species were recorded [34], bacteriocin of *L. fermentum* (10 kDa) had no effect against *S. aureus* and *Yersinia enterocolitica*. Recently Heredia-Castro et. al., [35] recorded that *Lactobacillus* species from cheese were shown to produce bacteriocin-like substances active against *S. aureus*, *L. innocua*, *E. coli* and *S. Typhimurium* by using the disk diffusion. Another study showed that bacteriocins of vaginal lactobacilli were steady at pH 4.5 - 7 but sensitive to pH 9 [36]. May this stability lead to make lactobacilli more effective on vaginal pathogens. In conclusion: The effectiveness of natural lactobacilli from vagina consider as good therapeutics against bacterial illness of humans. We need future study to purification the active component like bacteriocin or other organic acids from these bacteria.

References

1. Sieladie DV, Zambou NF, Kaktcham PM, Cresci A, Fonteh F. Probiotic properties of lactobacilli strains isolated from raw cow milk in the western highlands of Cameroon. *Innovative Romanian Food Biotechnology* 2011;9: 12-28.
2. Aasen IM, Mretr T, Katla T, Axelsson L, Storr I, et al. Influence of complex nutrients, temperature and pH on bacteriocin production by *Lactobacillus sakei* CCUG 42687. *Appl. Microbiol. Biotechnol* 2000;53 159-166.
3. Suskovic B, Kos J, Goreta S. Role of Lactic Acid Bacteria and Bifidobacteria in Synbiotic Effect. *Food Technol. Biotechnol* 2001;39:227-23.
4. Makarova K, Slesarev Wolf Y, Sorokin A, et al. Comparative genomics of the lactic acid bacteria". *Proc Natl Acad Sci U S A*. 2006;103(42):15611-6.
5. Goldstein E, Tyrrell L, Citron. *Lactobacillus* Species: Taxonomic Complexity and Controversial Susceptibilities. *clinical Infectious Diseases* 2015;60 (suppl. 2): S98-S107.
6. Mastromarino P, Vitali B, Mosca L. Bacterial vaginosis: a review on clinical trials with probiotics. *Hewmicrobiologica* 2013;36, 229-23.
7. McLean NW, Rosenstein IJ. Characterisation and selection of a *Lactobacillus* species to recolonise the vagina of women with recurrent bacterial vaginosis. *J Med Microbiol* 2000;49(6):543-452.
8. Forsum U, Hallén A, Larsson PG. Bacterial vaginosis a laboratory and clinical diagnostics enigma. *APMIS* 2005;113:153-61.
9. Hawes SE, Hillier SL, Benedetti J, et al. Hydrogen peroxide-producing lactobacilli and acquisition of vaginal infections. *J Infect Dis* 1996;174:1058-63.
10. Petrova I, Elke L, Shweta M, Nicole I, Lebeer S. "Lactobacillus species as biomarkers and agents that can promote various aspects of vaginal health". *Frontiers in Physiology* 2015;6.
11. Ma, Bing; Forney, Larry J.; Ravel, Jacques. "Vaginal Microbiome: Rethinking Health and Disease". *Annual Review of Microbiology* 2012;66 (1):371-389.
12. Helland MH, Wicklund T, Narvhus JA. Growth and metabolism of selected strains of probiotic bacteria, in maize porridge with added malted barley. *Int. J. Food. Microbiol* 2004;91:305-313.
13. Parvez S, Malik KA, Ah Kang S, Kim HY. Probiotics and their fermented food products are beneficial for health. *J. Appl. Microbiol* 2006;100: 1171-1185.
14. Jalilsood T, Baradaran A, Song AAL, Foo HL, Mustafa S, et al. Inhibition of pathogenic and spoilage bacteria by a novel biofilm forming *Lactobacillus* isolate: a potential host for the expression of heterologous proteins. *Microb .Cell. Fact* 2015;14: 283-288.
15. FAO-WHO. Food and agriculture organization of the United Nations,

- World Health Organization. Report of a joint FAO-WHO expert consultation on evaluation of health and nutritional properties of probiotics in food including powder milk with live lactic acid bacteria. Cordoba, Argentina 2001;2.
16. Manzoor A, Ul-Haq I, Baig S, Qazi, JI, Seratic, S. Efficacy of Locally Isolated Lactic Acid Bacteria Against Antibiotic-Resistant Uropathogens. *J Microbiol* 2016;9(1): e18952.
 17. Reid, G.; Bruce, A.W.; Fraser, N.; Heinemann, C.; Owen, J.; Henning, B. (2001). Oral probiotics can resolve urogenital infections. *FEMS Immunol. Med. Microbiol.*, 30, 49-52.
 18. Eschenbach, D.A.; Hillier, S.; Critchlow, C.; Stevens, C.; DeRouen, T.; Holmes, K.K. (1988). Diagnosis and clinical manifestations of bacterial vaginosis. *Am. J. Obstet. Gynecol.*, 158, 819-828.
 19. Larsson, P.G.; Forsum, U. Bacterial vaginosis - a disturbed bacterial flora and treatment enigma. *APMIS* 2010;113:305-316.
 20. Sobel J.D. Vulvovaginal candidiasis. *Lancet* 2007;369:1961-1971.
 21. Owen, M.K.; Clenney, T.L. Management of vaginitis. *Am. Fam. Physician* 2004;70:2125-2132.
 22. Paulitsch A, Weger W, Ginter-Hanselmayer G, Marth E, Buzina, W. A 5-year (2000-2004) epidemiological survey of *Candida* and non-*Candida* yeast species causing vulvovaginal candidiasis in Graz, Austria. *Mycoses* 2006;49: 471-475.
 23. Natalia F. Gil; Rafael C.R. Martinez; Bruna C. Gomes; Auro Nomizo; Elaine C. P. De Martinis. Vaginal lactobacilli as potential probiotics against *Candida* spp. *Braz. J. Microbiol* 2010; 41 (1):6-14.
 24. Zou J, Dong J, Yu X. Meta-analysis: Lactobacillus containing quadruple therapy versus standard triple first-line therapy for *Helicobacter pylori* eradication. *Helicobacter* 2009; 14:97-107.
 25. Zainab Farooq Shafeeq AL-Ryyis. Study the Effect of Bacteria *Lactobacillus* species on the Growth and Vitality of *Entamoeba histolytica* Isolated from Stool Samples of Patients. Msc.thesis. University of Baghdad. Mandal 201.
 26. Mandal SM. Coriander (*Coriandrum sativum* L.) essential oil: chemistry and biological activity. *Asian Pacific J. Trop Biomed* 2015;5: 421-428.
 27. Davoodabadi A, Dallal MMS, Lashani E, Ebrahimi MT. Antimicrobial activity of *Lactobacillus* spp. isolated from fecal flora of healthy breast-fed infants against diarrheagenic *Escherichia coli*. *Jundishapur J Microbiol* 2015;8: e27852.
 28. Ruhe JJ, Myers L, Mushatt D, Hasbun R. High-level penicillin-nonsusceptible *Streptococcus pneumoniae* bacteremia: identification of a low-risk subgroup. *Clin. Infect Dis* 2004;38:508.
 29. Vanderkooi OG, Low DE, Green K, et al. Predicting antimicrobial resistance in invasive pneumococcal infections. *Clin Infect Dis* 2005; 40:1288.

30. Subramanyam Dasari¹, Raju Naidu Devanaboyaina Shouri, Rajendra Wudayagiri, Lokanatha Valluru¹. Antimicrobial activity of *Lactobacillus* against microbial flora of cervicovaginal infections. *Asian Pac. J. Trop. Dis* 2014;4(1):18-24.
31. Jose NM, Bunt CR, Hussain MA. Implications of antibiotic resistance in probiotics. *Food Rev Int* 2015;31:52-62.
32. Strom K, Sjogren J, Broberg A, Schnürer J. *Lactobacillus Plantarum* MiLAB 393 produces the antifungal cyclic dipeptides cyclo(L-Phe-L-Pro) and cyclo (L-Phe-trans-4-OH-L-Pro) and 3-phenyllactic acid. *Appl. Environ. Microbiol* 2002;68:4322-4327.
33. Mohammed SD, Ijah UJJ. Isolation and screening of lactic acid bacteria from fermented milk products for bacteriocin production. *Annals Food Science and Technology* 2013;14:122-128.
34. Eid R, El Jakee J, Rashidy A, et al. Potential Antimicrobial Activities of Probiotic *Lactobacillus* Strains Isolated from Raw Milk. *J. Prob. Health* 2013; 4:2.
35. Heredia-Castro PY, Méndez-Romero JI, Hernández-Mendoza A, et al. Antimicrobial activity and partial characterization of bacteriocin-like inhibitory substances produced by *Lactobacillus* spp. isolated from artisanal Mexican cheese. *J.Dairy. Sci* 2015;98:8285-8293.
36. Alpay S, Aydin F, Kilich SS. Antimicrobial activity and characteristics of bacteriocins produced by vaginal *Lactobacilli*. *Turk J. Med. Sci* 2003;33: 7-13.

